



Hepatic and renal SOD and CAT gene expressions were determined using a semi quantitative RT-PCR according to [15]. Total RNA was prepared from the frozen hepatic and renal powder using the E.Z.N.A™ spin column RNA extraction kit (Omega Bio-Tech, Cat NO R6834-01, Canada) following the manufacturer instructions. Concentrations of RNA were measured by spectrophotometry (OD 260 nm), and RNA integrity was electrophoretic ally verified using ethidium bromide. After DNase treatment (Ambion, Clinsciences, Montrouge, France), RNA was reverse transcribed using Super Script II RNase H Reverse Transcriptase (Invitrogen, Carlsbad, CA, USA) in the presence of Random Primers (Promega, Charbonnières-les-Bains, France). Polymerase chain reaction (PCR) was performed using a 2720 thermocycler (Applied Biosystems, USA). Using PCR master mix (Qiagen USA) following the manufacturer instructions and using the specific primers for SOD "forward 5'-GGTGCCTGGAGCCCTA-3' and reverse 5'-ATCGGAAGTCTTCCACTGTC-3 and for CAT "forward 5'-TCCTGAATGAGGAGGAGCGA-3' and reverse 5'-ATCTTAGATGAGCGGTGATG -3', primer were designed using primer 3 programme based on the published nucleotide sequence information of the *O. niloticus* SOD and CAT genes (GenBank accession no. JF801727.1 and JF801726.1 ), PCR conditions were a denaturation at 95 °C for 2 min followed by 28 cycles of 95°C, 1 min; 55°C, 1 min; 72°C, 1 min . PCR products were analyzed on a 2% agarose gel in 90 mM Trisborate, 2 mM EDTA buffer (TBE), pH 8, and visualized by staining with ethidium bromide and UV transillumination, for quantitative evaluation, absolute optical densities (OD) of RT-PCR signals were obtained by densitometric scanning using an image analysis system (1-D Manager; TDI Ltd.). The values for the specific targets were normalized according to those of GAPDH to express arbitrary units of relative abundance of the specific messages (i.e., relative expression).

### 3. RESULTS

This work was designed to investigate the toxic effects of three different concentrations (1/3, 1/10 and 1/20 of LC50) of tea seed cake on *O. niloticus* through monitoring their effects on functional and histological of both liver and kidney, our results revealed that all used concentrations were toxic and the degree of toxicity related to the concentrations. The toxicity was manifested by increasing significantly ( $P \leq 0.05$ ;  $P \leq 0.01$ ) the concentrations of liver and kidney biochemical markers; serum ALT, creatinine and urea concentrations as well as the induction of the activities of the antioxidant enzymes; SOD and CAT. In both liver and kidney tissue (Table 1) on a molecular level there are induction of the gene expression of SOD and CAT genes in liver and kidney tissues (Figure 3). The histological examination of both kidney and liver tissues confirmed the biochemical and molecular results and revealed that, the hepatopancreatic tissue showed a severe vacuolization with pyknotic nuclei and intravascular hemolysis, necrosis with lymphocytes and EGCs infiltrations and hemosiderosis that stained blue by Prussian blue stain at the 1/3 LC50 concentration (Figure 1 B, C and D), but with 1/10 LC50; a moderate vacuolization in the hepatocytes and intravascular hemolysis were occurred (Figure 1 E) finally with 1/20 LC50; a centrolobular hydropic degeneration with pyknotic nuclei and mild intravascular hemolysis were manifested (Figure 1 F). In relation to kidney tissue it had a severe hydropic degeneration and vacuolization in the tubular epithelia with few round cells infiltration , focal coagulative necrosis and basophilic calcification that stained black by Von Kossa stain ( Figure 2 B,C and D), in 1/3 LC50, with 1/10 LC50 a vacuolization in the renal epithelia and intravascular hemolysis, lymphocytes aggregations around necrotic renal tubules were revealed (Figure 2 E and F), the same pathological manifestation were seen with 1/20 LC50 but in a low degree with thickening of tunica media ( Figure 2 G).

### 4. DISCUSSION

Tea seed cake is botanical pesticides could be extensively used in aquaculture to eliminate predatory fishes in fish and prawn ponds. It also widely used in killing snails in pond or coastal cropland, earthworms in vegetable field and underground pests in golf grassland. It can help shrimp exuviate and improve the quality of water. It not left contamination and poison through hydrolysing process in pond still crude protein and crude fibre, which can be assimilated thoroughly by aquatic creatures and algae bloom. Tea seed cake produced from *Camellia* sp. seeds after oil extraction it contains many active principles especially saponin which represent 5.2% - 7.2%. It has many toxicological effects on fish especially when be used in large concentrations. Our results indicated various levels of toxicity on *O. niloticus* fingerlings with respect to the different concentrations 1/3, 1/10 and 1/20 of LC50. On the serum level there was a high increase of liver and kidney toxicity biomarker ALT, creatinine and urea in fishes subjected to 1/3 LC50 (Table 1) , this may be due to the distraction

that occurred in liver and kidney cells as seen in histopathological examinations which revealed a severe vacuolization and necrosis of hepatocyte (Figure 1B, C and D), Kidney shows severe hydropic degeneration and vacuolization in the renal tubular epithelia and few round cells infiltration, focal coagulative necrosis and basophilic calcification (Figure 2 B,C and D) the concentration of the biochemical markers (ALT, creatinine and urea) were decreased gradually thorough decreasing of tea seed cake concentration 1/10 and 1/20 LC50 ( Table 2) but still significantly increased when compared with control samples, the improvement of serum hepatic and renal biomarkers concentration may be due to the moderate effect of tea seed cake on hepatocyt which manifested by moderate vacuolization and degeneration (Figure 1 E and F), Kidney shows moderate vacuolization and necrosis in the renal epithelia (Figure 2 E, F and G). The toxic effect of tea seed cake may be explained by its high level of saponin. The toxic effect of saponin of *Quillaja saponaria* that manifested by the increase of serum ALT and the histological observations showed hepatocyte injures, hypertrophy, cloudy swelling and vacuolization with architectural disarray, hepatic cell degeneration, kidney tubule necrosis with haemolytic erythrocytes in the hematopoietic tissue appeared [16]. In rat saponins mixture revealed mild cytolysis of the hepatocytes affecting predominantly the periportal region, nuclear vacuolation and prominent nuclei and sinusoidal congestion hepatocytes architectural disarray with extensive necrosis, kidney showed focal tubular atrophy and necrosis with epithelial cells vacuolation and tubular necrosis other investigators have reported untoward histopathological changes in rat /mice tissues following ingestion of saponin containing substances changes observed in transaminases activities in liver and plasma of rats was speculated to be a reflection of liver damage occasioned by ingestion of saponin while the increase of plasma creatinine and urea might be due to kidney damage [17-20]. Mice ingested saponins had a hepatotoxic lesion that manifested by small hemorrhage in many hepatic lobules, congestion of central veins and sinusoid, distraction of liver architecture and hepatic necrosis as well destruction of renal tubular cells and haemorrhage [21]. CAT gene expression and activities in liver and kidney tissues of fish exposed to tea seed cake were found to be higher than in control fish (Table 1 and figure 3). The stimulation of CAT gene expression and activities were clearly concentration dependant, it increased in response to 1/10 and 1/20 LC50 of toxin but the increase was most pronounced in fish exposed to the highest concentration of toxin (1/3LC50). CAT is an enzyme that is known to detoxify reactive oxygen species in the cells. The increase of gene expression and activity of the CAT in fish in response to tea seed cake most probably occurs in response to an increase of reactive oxygen species indicating an oxidative stress in the animal in response to the toxin. The hepatic and renal SOD activities and gene expression were higher in fish exposed to tea seed cake than in control fish (Table 1 and figure 3) this increase is a concentration dependant. The increase in SOD activities and gene expression may be explained as a response to the toxic effect of tea seed cake on both liver and kidney tissues which may be induced the super oxide radical's generation. As with other animals, fish have antioxidant defense mechanisms, which help to maintain health and prevent oxidation lesions. SOD and CAT are important antioxidant enzymes [22-23]. The enzymes are commonly used in toxicological tests as stress indicators [24]. SOD and CAT are scavengers of the reactive oxygen species, acting on superoxide (O<sub>2</sub><sup>-</sup>) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) respectively [23].

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